

# S<sub>0</sub>-State Model of the Oxygen-Evolving Complex of Photosystem II

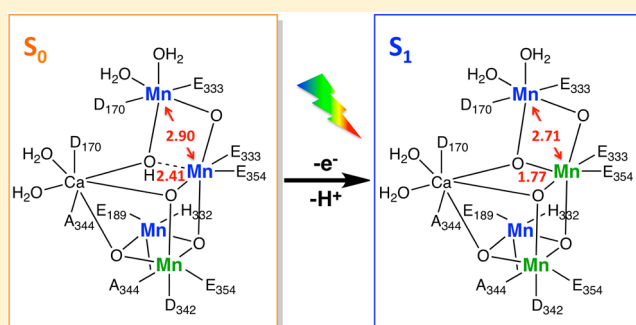
Rhitankar Pal,<sup>†</sup> Christian F. A. Negre,<sup>†</sup> Leslie Vogt,<sup>†</sup> Ravi Pokhrel,<sup>†</sup> Mehmed Z. Ertem,<sup>†,‡</sup> Gary W. Brudvig,<sup>\*,†</sup> and Victor S. Batista<sup>\*,†</sup>

<sup>†</sup>Department of Chemistry, Yale University, New Haven, Connecticut 06511, United States

<sup>‡</sup>Department of Chemistry, Brookhaven National Laboratory, Building 555A, Upton, New York 11973, United States

## Supporting Information

**ABSTRACT:** The S<sub>0</sub> → S<sub>1</sub> transition of the oxygen-evolving complex (OEC) of photosystem II is one of the least understood steps in the Kok cycle of water splitting. We introduce a quantum mechanics/molecular mechanics (QM/MM) model of the S<sub>0</sub> state that is consistent with extended X-ray absorption fine structure spectroscopy and X-ray diffraction data. In conjunction with the QM/MM model of the S<sub>1</sub> state, we address the proton-coupled electron-transfer (PCET) process that occurs during the S<sub>0</sub> → S<sub>1</sub> transition, where oxidation of a Mn center and deprotonation of a μ-oxo bridge lead to a significant rearrangement in the OEC. A hydrogen bonding network, linking the D1-D61 residue to a Mn-bound water molecule, is proposed to facilitate the PCET mechanism.



Photosystem II (PSII) is a 650 kDa protein complex found in the thylakoid membranes of higher plants, algae, and internal membranes of cyanobacteria. It is the only biological machinery that utilizes solar energy for water oxidation,<sup>1–4</sup> extracting reducing equivalents from water and generating molecular O<sub>2</sub> as a byproduct. The PSII complex consists of ~20 protein subunits, numerous electron-transport cofactors, chlorophyll and β-carotene pigments, and a CaMn<sub>4</sub>O<sub>5</sub> catalytic core, known as the oxygen-evolving complex (OEC).<sup>5,6</sup> In PSII, absorption of a photon triggers an efficient charge separation across the membrane bilayer, stabilizing the generation of a radical cation on a pair of chlorophylls, P<sub>680</sub><sup>•+</sup>. In the charge-separated state, P<sub>680</sub><sup>•+</sup>, with an estimated E° of 1.25 V, is the primary oxidant in PSII. In each step of the catalytic cycle, P<sub>680</sub><sup>•+</sup> oxidizes a redox-active tyrosine residue (Y<sub>Z</sub>), which in turn oxidizes the OEC. Four successive photo-oxidation steps are required for a complete cycle turnover, advancing the redox state of the OEC through five storage states (S states, S<sub>i</sub>, where i = 0–4). The S<sub>0</sub> state is the most reduced state, and four sequential oxidations transform the OEC into the S<sub>4</sub> state. An O–O bond forms in the S<sub>4</sub> state, upon which the OEC relaxes back to the S<sub>0</sub> state. All but the S<sub>1</sub> → S<sub>2</sub> oxidative transition involve the release of a proton to the lumen. Hence, efficient transfer of a proton from the OEC to the lumen is critical for redox leveling during successive oxidation steps in the S-state cycle. Therefore, elucidating the redox leveling mechanism in the oxidation of water at the OEC is of fundamental importance for the design of artificial systems capable of catalyzing the water oxidation reaction with a similar efficiency.

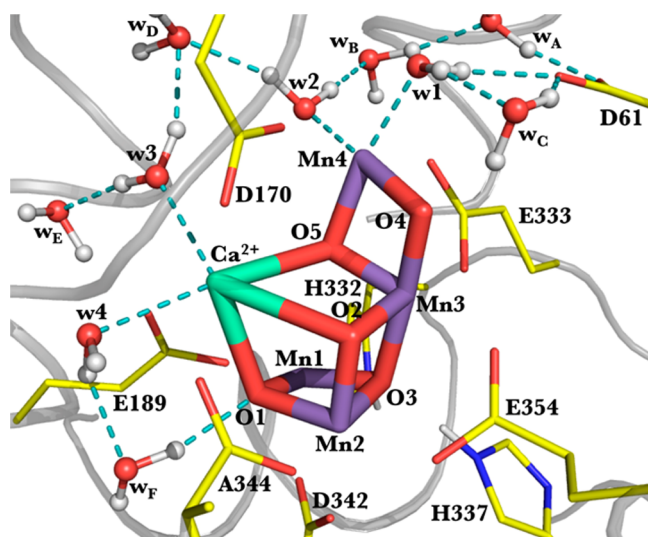
Biophysical studies based on time-resolved mass spectrometry,<sup>7–9</sup> electron paramagnetic resonance (EPR) spectroscopy,<sup>10–12</sup> Fourier transform infrared (FTIR) spectroscopy,<sup>13–15</sup>

and X-ray diffraction (XRD),<sup>16–20</sup> as well as computational modeling,<sup>21–28</sup> have been instrumental in gaining structural and mechanistic insights. In particular, multiple research groups over the past decade have reported XRD models of PSII at progressively higher resolution (3.8–1.9 Å).<sup>16–20</sup> These breakthroughs in XRD have converged into a model of the OEC that involves a cuboidal CaMn<sub>3</sub> cluster with a dangling Mn held together by putative μ-oxo bridges and protein side chains. The model is consistent with earlier proposals based on EPR and extended X-ray absorption fine structure (EXAFS) spectroscopy.<sup>11,29,30</sup> The XRD model at 1.9 Å resolution has resolved for the first time terminal water molecules directly bound to Ca<sup>2+</sup> and the dangling Mn.<sup>20</sup>

Several new features, including an additional μ-oxo bridge connecting the dangling Mn with the CaMn<sub>3</sub>O<sub>4</sub> cube and D1-D170 as a bridging ligand between Ca<sup>2+</sup> and the dangling Mn, were also resolved in the 1.9 Å XRD structure.<sup>20</sup> Furthermore, carboxylate ligands D1-E333, CP43-E354, and D1-D342 and the C-terminus of D1-A344 were shown to bridge the Mn and Ca<sup>2+</sup> ions of the cluster (see Figure 1). The D1-E189 and D1-H332 side chains were shown to coordinate Mn1 directly. However, simulations of EXAFS spectra suggested that the XRD model of the OEC represents an average structure of the OEC in multiple oxidation states, including the states of the OEC more reduced than those present in the catalytic cycle of water splitting.<sup>24,31</sup> The XRD model has allowed the development of a quantum mechanics/molecular mechanics (QM/MM) model of the OEC

**Received:** September 3, 2013

**Revised:** October 10, 2013



**Figure 1.** Quantum mechanics/molecular mechanics-optimized  $S_1$ -state structure of the OEC  $\text{CaMn}_4\text{O}_5$  cluster, along with all directly ligated amino acid residues (D170, E189, H332, E333, H337, D342, A344, and E354), residue D61, and 10 water molecules forming crucial hydrogen bonding networks. Four water molecules are directly ligated to the OEC ( $w_1$ – $w_4$ ), and six others (labeled  $w_A$ – $w_F$ ) are in the second sphere.

in the  $S_1$   $\text{Mn}_4[\text{III,IV,IV,III}]$  state that is consistent with X-ray data, showing that  $\text{Ca}^{2+}$  is bridged to Mn by the carboxylate moieties of D170 and A344, and with high-resolution spectroscopy, including polarized EXAFS data of oriented single crystals.<sup>24</sup> While several density functional theory (DFT) models for the  $S_0$  state have been suggested,<sup>21–23</sup> a structural model that is fully consistent with EXAFS and XRD data has yet to be reported. In conjunction with the previously reported QM/MM  $S_1$  model,<sup>24</sup> the  $S_0$  structure would provide a mechanistic proposal for the  $S_0 \rightarrow S_1$  transition in which oxidation of a Mn ion and deprotonation of a  $\mu$ -hydroxo bridge lead to significant rearrangements in the OEC.

Using QM/MM optimizations followed by Monte Carlo (MC) refinements, as in our prior work, we propose an  $S_0$  model, which suggests that the  $S_0 \rightarrow S_1$  PCET mechanism could involve deprotonation of the O5  $\mu$ -hydroxo bridge via hydrogen-bonded water molecules linking Mn4 and the D1–D61 residue. Furthermore, we demonstrate that the structural models obtained with this approach for the  $S_1$  and  $S_0$  states are consistent with both EXAFS<sup>29</sup> and XRD<sup>20</sup> data.

The QM/MM models were built using a two-layer ONIOM link H-atom approach implemented in Gaussian09,<sup>32</sup> as in our previous studies.<sup>24,26,27</sup> However, in this work, we expanded the QM region to move the QM/MM boundary farther from the OEC and to better model the hydrogen bonding network surrounding the cluster. The  $S_1$  model has been reoptimized with this expanded QM region for the sake of comparison. The QM layer includes the OEC, all directly ligated side chains (D1–D170, D1–E189, D1–H332, D1–E333, D1–D342, and CP43–E354), the C-terminus of D1–A344, hydrogen-bonded residues D1–H337 and CP43–R357, residue D1–D61, and 10 water molecules shown in Figure 1 (further details are provided in section I of the Supporting Information). The MM layer includes all residues with  $\alpha$  atoms within 15 Å of the OEC, crystallographic waters within this boundary, two chloride ions, and amino acids that span gaps of up to two residues in the peptide chains of the main selection (section I of the Supporting Information). QM/MM

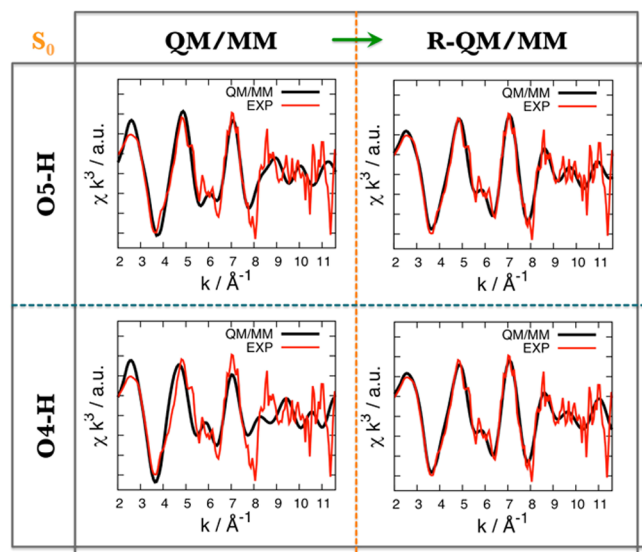
optimizations were performed using the B3LYP<sup>33,34</sup> functional with the LANL2DZ<sup>35,36</sup> pseudopotential for Ca and Mn and the 6-31G\*<sup>37</sup> basis set for all other atoms. The AMBER force field<sup>38</sup> was used for all MM layer atoms. EXAFS spectra were calculated using the *ab initio* real space Green's function approach as implemented in FEFF (version 8.30).<sup>39</sup> EXAFS structural refinement was based on simulated annealing MC (see section I of the Supporting Information), fully relaxing the geometry of the  $\text{CaMn}_4\text{O}_5$  cluster, and relaxing all the directly coordinated ligands with harmonic constraints relative to the QM/MM-optimized positions (R-QM/MM).

The calculated EXAFS for the reoptimized QM/MM  $S_1$  model agrees very well with the experimental spectrum (Figure S3 of the Supporting Information). It is important to note that the optimization of the isolated DFT model layer leads to a worse match for the EXAFS spectrum and requires larger displacements in the MC refinement (section II and Figure S4 of the Supporting Information). Therefore, the advantage of performing QM/MM optimization rather than using a DFT cluster model is clearly reflected in the better agreement with the experimental EXAFS spectrum, which is also consistent with the recent findings of Retegan et al.<sup>28</sup>

Having a reasonable structure of the  $S_1$  state using QM/MM optimization, we used this approach to investigate the structure of the  $S_0$  state. To determine the likely site for protonation of the OEC in the  $S_0$  state, a preliminary MC screening using a simulated annealing algorithm<sup>40</sup> was performed on the geometry of the  $S_1$  model to fit the  $S_0$  EXAFS data. MC optimization revealed that the Mn3, Mn4, O4, and O5 atoms of the OEC had significant displacements, thereby suggesting possible protonation at the O5 or O4  $\mu$ -oxo bridges in the  $S_0$  state (see the Supporting Information for further details). Furthermore, this also indicated that either Mn3 or Mn4 must be oxidized during this transition to induce the requisite geometry change. These results are in agreement with the reported  $B$  factors of the atoms in the X-ray crystal structure at 1.9 Å resolution (Protein Data Bank entry 3ARC).<sup>20</sup> The other  $\mu$ -oxo atoms are unlikely to be protonated as O2 is near the positively charged R357 and O3 is strongly hydrogen bonded to H337. In light of these observations, we explored the protonation of O4 and O5 sites using our QM/MM model of the  $S_0$  state. The EXAFS spectra of the O4–H and O5–H models (QM/MM and R-QM/MM) are shown in Figure 2. An overlay of the QM/MM and R-QM/MM structures of the  $S_0$  state where either O5 or O4 is protonated is shown in Figure S6 (section IV of the Supporting Information).

The intermetallic distances listed in Table 1 for the  $S_0$  and  $S_1$  QM/MM models provide a clear description of the underlying structural rearrangements induced by the  $S_0 \rightarrow S_1$  oxidation-state transition. In the  $S_0$  state, the formal oxidation number of Mn3 is III, and upon oxidation to the  $S_1$  state, the formal oxidation number of Mn3 increases to IV. This shortens the Mn3–O5 distance, inducing deprotonation of the O5–H  $\mu$ -hydroxo bridge and shortening the Mn3–Mn4 distance from 2.9 to 2.7 Å (Figure 3, top panel). As a result, the  $S_1$  model has three short Mn–Mn distances of  $\sim 2.7$  Å (Mn1–Mn2, Mn2–Mn3, and Mn3–Mn4) and a long Mn1–Mn3 distance of  $\sim 3.2$  Å.

In contrast, the  $S_0$  model has only two short Mn–Mn distances of  $\sim 2.7$  Å (Mn1–Mn2 and Mn2–Mn3), while the other two Mn–Mn distances (i.e., Mn1–Mn3 and Mn3–Mn4) are longer ( $\sim 2.9$ – $3.0$  Å). The elongated Mn3–Mn4 distance in the  $S_0$  state is the major difference between the  $S_0$  and  $S_1$  models (Figure 3) and is clearly manifested in the comparison of Fourier transform EXAFS spectra (Figure 3, bottom panel). In the 2.0–3.0 Å region



**Figure 2.** Comparison of the experimental EXAFS spectrum for the OEC  $S_0$  state<sup>29</sup> and the calculated spectra based on the DFT-QM/MM (left) and R-QM/MM (right) models with  $\mu$ -hydroxo bridges at O5 (top) and O4 (bottom).

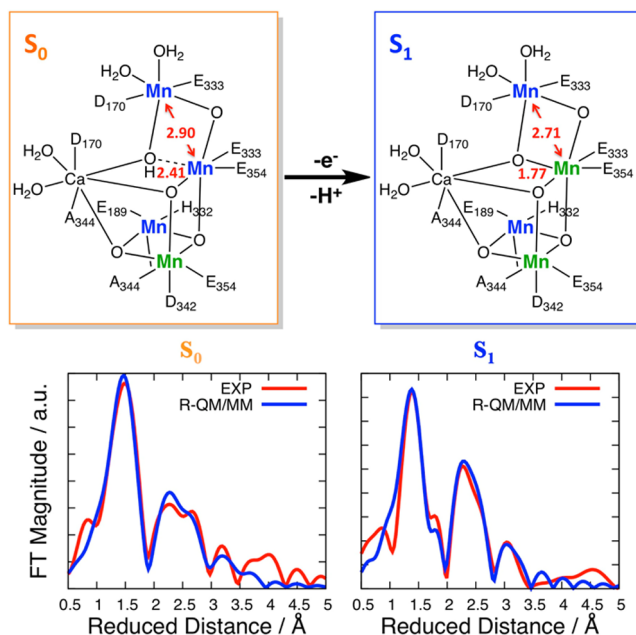
**Table 1. Intermetallic Distances (in angstroms) in the X-ray Structure<sup>20</sup> and in the R-QM/MM Models of the  $S_0$  (O5 protonated) and  $S_1$  States**

	X-ray	$S_1$	$S_0$ O5-H
	chain A/a	QM/MM (R-QMMM)	QM/MM (R-QMMM)
Mn1–Mn2	2.84/2.76	2.73 (2.74)	2.72 (2.74)
Mn1–Mn3	3.29/3.30	3.26 (3.27)	3.20 (3.19)
Mn1–Mn4	5.00/4.95	4.76 (4.70)	4.72 (4.66)
Mn2–Mn3	2.89/2.91	2.76 (2.75)	2.77 (2.76)
Mn2–Mn4	5.44/5.37	5.02 (5.02)	5.25 (5.19)
Mn3–Mn4	2.97/2.91	2.68 (2.71)	2.95 (2.90)
Ca <sup>2+</sup> –Mn1	3.51/3.46	3.55 (3.52)	3.37 (3.41)
Ca <sup>2+</sup> –Mn2	3.36/3.29	3.44 (3.50)	3.46 (3.45)
Ca <sup>2+</sup> –Mn3	3.41/3.44	3.58 (3.65)	3.72 (3.73)
Ca <sup>2+</sup> –Mn4	3.79/3.80	3.68 (3.70)	3.92 (3.91)

of reduced distance, a single peak in the  $S_1$  state splits into a doublet in the  $S_0$  state (Figure 3, bottom panel, and section VI of the Supporting Information).

Recent experimental observations suggest that the oxidation of the OEC precedes the release of the proton to the lumen, during the  $S_0 \rightarrow S_1$  transition, unlike the later S-state transitions in which proton release occurs prior to oxidation of the OEC.<sup>41</sup> However, the detailed mechanism responsible for the translocation of the proton to the lumen has yet to be established.

We have analyzed the possibility of a Grotthuss-like mechanism<sup>42</sup> for deprotonation during the  $S_0 \rightarrow S_1$  transition in which D1-D61 is involved in the transfer of the proton to the lumen.<sup>26,27,43</sup> We examined the removal of the proton from the O5–H hydroxo bridge, via the  $w_2 \rightarrow w_B \rightarrow w_A \rightarrow D61$  pathway. In the QM/MM  $S_0$  model, the hydroxo bridge of O5 does not form a direct hydrogen bond with any of the OEC-bound or second-shell water molecules. However, we find that a water ligand of the dangling Mn ( $w_2$ ) could function as a proton acceptor after deprotonation (Figure S7 of the Supporting Information). To test this hypothesis, an  $S_0'$  model was constructed by oxidizing the Mn3 center (III  $\rightarrow$  IV) and



**Figure 3.** Structural changes induced by the  $S_0 \rightarrow S_1$  transition (top). The Mn3–Mn4 and Mn3–O5 distances are shortened upon oxidation of Mn and deprotonation of the hydroxo bridge: green for Mn<sup>IV</sup> and blue for Mn<sup>III</sup>. Comparison of the experimental Fourier transform EXAFS spectra (red) and the calculated (blue) spectra of the  $S_0$  (left) and  $S_1$  (right) QM/MM models (bottom).

deprotonating the  $w_2$  molecule (Figure S7b of the Supporting Information), which can be facilitated by the hydrogen bonding network connecting  $w_2$  to the D1-D61 side chain. The DFT calculations indicate that the hydroxo ligand of Mn4 could deprotonate the O5–H  $\mu$ -hydroxo bridge through a barrierless and thermodynamically spontaneous process. The energetics of removal of the proton from O5–H to  $w_2(\text{OH}^-)$  was computed by increasing the O5–H bond length from 1.1 to 2.3 Å and relaxing the entire model at every step. The overall path was found to be exothermic by  $\sim 17$  kcal/mol (see section VI of the Supporting Information for details).

In summary, we conclude that the  $S_0 \rightarrow S_1$  transition of the OEC of PSII involves oxidation of Mn3 (from III to IV) and deprotonation of the O5–H  $\mu$ -hydroxo bridge. The oxidation reduces the Mn3–Mn4 intermetallic distance from 2.9 to 2.7 Å because of the removal of the Jahn–Teller distortion. This structural/oxidation-state transition strengthens the Mn3–O5 bond, leading to deprotonation of the OH ligand. The resulting structural rearrangements of the Mn cluster are fully consistent with the EXAFS spectroscopy of the  $S_0$ - and  $S_1$ -state intermediates. Deprotonation of the O5–H  $\mu$ -hydroxo bridge is likely mediated by a Grotthuss-like proton-transport mechanism that involves a stable hydrogen bonding network linking  $w_2$  and D1-D61 via bound water molecules  $w_B$  and  $w_A$ .

## ■ ASSOCIATED CONTENT

### ● Supporting Information

Description of computational methods, pictorial representation of the X-ray structure  $\beta$  factors on the atoms of the metal cluster at the center of the OEC, combined MC-DFT-generated EXAFS, and coordinates of the QM/MM and MC-refined models. This material is available free of charge via the Internet at <http://pubs.acs.org>.



## AUTHOR INFORMATION

### Corresponding Authors

\*E-mail: victor.batista@yale.edu. Phone: (203) 432-6672. Fax: (203) 432 6144.

\*E-mail: gary.brudvig@yale.edu. Phone: (203) 432-5202. Fax: (203) 432-6144.

### Funding

### Notes

The authors declare no competing financial interest.

## ACKNOWLEDGMENTS

V.S.B. acknowledges supercomputer time from NERSC and financial support from the Division of Chemical Sciences, Geosciences, and Biosciences, Office of Basic Energy Sciences, U.S. Department of Energy (DESC0001423). Biochemical work was supported by the Department of Energy, Office of Basic Energy Sciences, Division of Chemical Sciences (Grant DE-FG02-05ER15646 to G.W.B.). M.Z.E. was funded by a Computational Materials and Chemical Sciences (CMCSN) project at Brookhaven National Laboratory under contract DE-AC02-98CH10886 with the U.S. DOE and supported by its Division of Chemical Sciences, Geosciences & Biosciences, Office of Basic Energy Sciences.

## REFERENCES

- (1) Debus, R. J. (1992) *Biochim. Biophys. Acta* 1102, 269–352.
- (2) Dau, H., and Haumann, M. (2008) *Coord. Chem. Rev.* 252, 273–295.
- (3) Lubitz, W., Reijerse, E. J., and Messinger, J. (2008) *Energy Environ. Sci.* 1, 15–31.
- (4) Vinyard, D. J., Ananyev, G. M., and Dismukes, G. C. (2013) *Annu. Rev. Biochem.* 82, 577–606.
- (5) Kashino, Y., Lauber, W. M., Carroll, J. A., Wang, Q., Whitmarsh, J., Satoh, K., and Pakrasi, H. B. (2002) *Biochemistry* 41, 8004–8012.
- (6) McEvoy, J. P., and Brudvig, G. W. (2006) *Chem. Rev.* 106, 4455–4483.
- (7) Messinger, J., Badger, M., and Wydrzynski, T. (1995) *Proc. Natl. Acad. Sci. U.S.A.* 92, 3209–3213.
- (8) Hillier, W., and Wydrzynski, T. (2004) *Phys. Chem. Chem. Phys.* 6, 4882–4889.
- (9) Hillier, W., and Wydrzynski, T. (2008) *Coord. Chem. Rev.* 252, 306–317.
- (10) Peloquin, J. M., Campbell, K. A., Randall, D. W., Evanchik, M. A., Pecoraro, V. L., Armstrong, W. H., and Britt, R. D. (2000) *J. Am. Chem. Soc.* 122, 10926–10942.
- (11) Britt, R. D., Campbell, K. A., Peloquin, J. M., Gilchrist, M. L., Aznar, C. P., Dicus, M. M., Robblee, J., and Messinger, J. (2004) *Biochim. Biophys. Acta* 1655, 158–171.
- (12) Kulik, L. V., Epel, B., Lubitz, W., and Messinger, J. (2005) *J. Am. Chem. Soc.* 127, 2392–2393.
- (13) Chu, H. A., Hillier, W., and Debus, R. J. (2004) *Biochemistry* 43, 3152–3166.
- (14) Debus, R. J. (2008) *Coord. Chem. Rev.* 252, 244–258.
- (15) Noguchi, T. (2008) *Coord. Chem. Rev.* 252, 336–346.
- (16) Zouni, A., Witt, H. T., Kern, J., Fromme, P., Krauss, N., Saenger, W., and Orth, P. (2001) *Nature* 409, 739–743.
- (17) Ferreira, K. N., Iverson, T. M., Maghlaoui, K., Barber, J., and Iwata, S. (2004) *Science* 303, 1831–1838.
- (18) Loll, B., Kern, J., Saenger, W., Zouni, A., and Biesiadka, J. (2005) *Nature* 438, 1040–1044.
- (19) Guskov, A., Kern, J., Gabdulkhakov, A., Broser, M., Zouni, A., and Saenger, W. (2009) *Nat. Struct. Mol. Biol.* 16, 334–342.
- (20) Umena, Y., Kawakami, K., Shen, J. R., and Kamiya, N. (2011) *Nature* 473, 55–60.

- (21) Isobe, H., Shoji, M., Yamanaka, S., Umena, Y., Kawakami, K., Kamiya, N., Shen, J. R., and Yamaguchi, K. (2012) *Dalton Trans.* 41, 13727–13740.
- (22) Siegbahn, P. E. (2013) *Biochim. Biophys. Acta* 1827, 1003–1019.
- (23) Ichino, T., and Yoshioka, Y. (2012) *Chem. Phys. Lett.* 545, 107–111.
- (24) Lubner, S., Rivalta, I., Umena, Y., Kawakami, K., Shen, J. R., Kamiya, N., Brudvig, G. W., and Batista, V. S. (2011) *Biochemistry* 50, 6308–6311.
- (25) Sproviero, E. M., Gascon, J. A., McEvoy, J. P., Brudvig, G. W., and Batista, V. S. (2006) *J. Chem. Theory Comput.* 2, 1119–1134.
- (26) Sproviero, E. M., Gascon, J. A., McEvoy, J. P., Brudvig, G. W., and Batista, V. S. (2008) *J. Am. Chem. Soc.* 130, 3428–3442.
- (27) Sproviero, E. M., Gascon, J. A., McEvoy, J. P., Brudvig, G. W., and Batista, V. S. (2008) *J. Am. Chem. Soc.* 130, 6728–6730.
- (28) Retegan, M., Neese, F., and Pantazis, D. A. (2013) *J. Chem. Theory Comput.* 9, 3832–3842.
- (29) Haumann, M., Muller, C., Liebisch, P., Iuzzolino, L., Dittmer, J., Grabolle, M., Neisius, T., Meyer-Klaucke, W., and Dau, H. (2005) *Biochemistry* 44, 1894–1908.
- (30) Grabolle, M., Haumann, M., Muller, C., Liebisch, P., and Dau, H. (2006) *J. Biol. Chem.* 281, 4580–4588.
- (31) Galstyan, A., Robertazzi, A., and Knapp, E. W. (2012) *J. Am. Chem. Soc.* 134, 7442–7449.
- (32) Frisch, M. J., et al. (2009) *Gaussian 09*, Gaussian, Inc., Wallingford, CT.
- (33) Becke, A. D. (1988) *Phys. Rev. A* 38, 3098–3100.
- (34) Becke, A. D. (1993) *J. Chem. Phys.* 98, S648–S652.
- (35) Wadt, W. R., and Hay, P. J. (1985) *J. Chem. Phys.* 82, 284–298.
- (36) Hay, P. J., and Wadt, W. R. (1985) *J. Chem. Phys.* 82, 299–310.
- (37) Hariharan, P. C., and Pople, J. A. (1973) *Theor. Chim. Acta* 28, 213–222.
- (38) Case, D. A., et al. (2012) *AMBER 12*, University of California, San Francisco.
- (39) Ankudinov, A. L., Bouldin, C. E., Rehr, J. J., Sims, J., and Hung, H. (2002) *Phys. Rev. B* 65, 104107.
- (40) Kirkpatrick, S., Gelatt, C. D., and Vecchi, M. P. (1983) *Science* 220, 671–680.
- (41) Klaus, A., Haumann, M., and Dau, H. (2012) *Proc. Natl. Acad. Sci. U.S.A.* 109, 16035–16040.
- (42) Cukierman, S. (2006) *Biochim. Biophys. Acta* 1757, 876–885.
- (43) Service, R. J., Hillier, W., and Debus, R. J. (2010) *Biochemistry* 49, 6655–6669.

## NOTE ADDED AFTER ASAP PUBLICATION

This paper published ASAP on October 17, 2013. The Supporting Information file was replaced and the revised version was reposted on October 21, 2013.